



Designation: D4445 – 23

Standard Test Method for Fungicides for Controlling Sapstain and Mold on Unseasoned Lumber (Laboratory Method)¹

This standard is issued under the fixed designation D4445; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This (laboratory) test method is used for determining the minimum concentration of fungicide, or formulation of fungicides, that is effective in preventing biodeterioration by sapstain fungi and molds in selected species of wood under optimum laboratory conditions.

NOTE 1—From the results of this test, commercial treating solution concentrations cannot be estimated without further field tests.

1.2 The requirements for test materials and procedures are discussed in the following order:

	Section
Summary of Test Method	4
Apparatus	6
Reagents	7
Wood	8
Test Fungi	9
Culture Media	10
Preparation of Inoculum	11
Preparation of Test Chambers	12
Treatment of Samples	13
Inoculation and Incubation	14
Evaluation of the Test	15
Report	16

1.3 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.4 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety, health, and environmental practices and determine the applicability of regulatory limitations prior to use.*

1.5 *This international standard was developed in accordance with internationally recognized principles on standardization established in the Decision on Principles for the Development of International Standards, Guides and Recommendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.*

¹ This test method is under the jurisdiction of ASTM Committee D07 on Wood and is the direct responsibility of Subcommittee D07.06 on Treatments for Wood Products.

Current edition approved April 15, 2023. Published May 2023. Originally approved in 1984. Last previous edition approved in 2019 as D4445 – 10 (2019). DOI: 10.1520/D4445-23.

2. Referenced Documents

2.1 ASTM Standards:²

- D9 Terminology Relating to Wood and Wood-Based Products
- D1165 Nomenclature of Commercial Hardwoods and Softwoods
- D1193 Specification for Reagent Water
- D5590 Test Method for Determining the Resistance of Paint Films and Related Coatings to Fungal Defacement by Accelerated Four-Week Agar Plate Assay

3. Terminology

3.1 *Definitions*—For definitions of terms used in this test method, refer to Terminologies D9 and D1165.

4. Summary of Test Method

4.1 Unseasoned sapwood specimens are treated either by spraying with, or by immersing in, solutions or dispersions of a fungicide formulation prepared at five or more concentration levels. The specimens are exposed to sapstain fungi and molds. Options for testing the toxicity of fungicides include testing against individual fungi or against several fungi by using a mixed spore suspension for the inoculation of the specimens.

4.2 The intensity of surface fungal growth is estimated after incubation and the results used to determine the minimum chemical treatment concentration giving zero growth (CGo).

5. Significance and Use

5.1 This test method is useful as a screening procedure for selecting fungicides or formulations for more rigorous field evaluation.

6. Apparatus

6.1 *Incubation Room (or Incubation Cabinet)*, maintained at a temperature of 25 °C ± 1 °C, and relative humidity between 70 % and 80 %.

6.2 *Steam Sterilizer.*

² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

6.3 Containers:

6.3.1 *Sterile Petri Dishes*, with minimum size of 140 mm (diameter) by 20 mm (height) with lid or,

6.3.2 *Aluminum Pans*, with minimum size of 240 mm by 100 mm by 20 mm (height) with aluminum foil cover.

6.4 Spacers:

6.4.1 *U-Shaped or Straight Glass Rods*, with 3 mm diameter or,

6.4.2 *Polyethylene Mesh*, cut to cover the bottom of the selected container(s).

7. Reagents

7.1 *Purity of Water*—Reference to water shall be understood to mean sterile reagent water conforming to Type IV of Specification D1193.

8. Wood

8.1 *General Properties*—The wood species to be tested shall be selected on the basis of their susceptibility to staining/mold fungi (pine or spruce species are preferred when evaluating softwoods). Sapwood of the selected wood species, unseasoned (moisture content higher than 40 %), free of knots, visible decay, sapstain, and mold, shall be used (Note 2). If the fungicide is to be used to protect hardwood, the inclusion of sapwood from a susceptible hardwood species, such as red oak or hickory, is recommended.

NOTE 2—If wood for the test is collected in a sawmill where logs are stored in water, it is necessary to collect lumber from at least three different logs since depletion of nutrients during water storage may strongly affect the growth of molds and staining fungi. Ensure that the lumber collected in a sawmill has not been treated with a sapstain and mold preventive, and if there is any doubt, at least 10 mm of surface wood must be removed and discarded.

8.2 *Size of Specimens*—Specimens shall be 7 mm by 20 mm in cross section and 70 mm long.

8.3 *Preparation of Specimens*—Within two days of collecting, the samples shall be cut from the wood using a sharp saw blade. To prevent drying, the specimens shall be stored in polyethylene bags. For storage longer than one day, but less than one year, tightly packed specimens shall be frozen (–20 °C or lower) in polyethylene bags. For these longer storage cases, the contents of one bag shall be limited to as many specimens as are used for a single experiment.

9. Test Fungi³

9.1 Hardwoods:

9.1.1 Sapstain Fungi:

9.1.1.1 *Diplodia gossypina* Cooke (Pole-Evans, anamorph) (ATCC 34643).

9.1.1.2 *Davidsoniella virescens* (Davison) de Beer et al., a form of *C. coerulescens* found on American hardwoods.

9.1.1.3 *Aureobasidium pullulans* (de Bary) Arnaud (ATCC 16624).

9.1.2 Mold Fungi:

9.1.2.1 *Trichoderma pseudokoningii* Rifai (ATCC 26801).

9.1.2.2 *Cephalosporium fragrans* Hanawa (ATCC 12091).

9.1.2.3 *Clonostachys rosea* (Link:Fries) (ATCC 10521).

9.2 Softwoods:

9.2.1 Sapstain Fungi:

9.2.1.1 *Diplodia gossypina* Cooke (Pole-Evans, anamorph) (ATCC 34643).

9.2.1.2 *Ceratocystis pilifera* (Fr.) C. Moreau (ATCC 15457).

9.2.1.3 *Aureobasidium pullulans* (de Bary) Arnaud (ATCC 16624).

9.2.2 Mold Fungi:

9.2.2.1 *Trichoderma pseudokoningii* (Rifai) (ATCC 26801).

9.2.2.2 *Cephalosporium fragrans* Hanawa (ATCC 12091).

9.2.2.3 *Clonostachys rosea* (Link:Fries) (ATCC 10521).

9.3 *General Consideration*—In addition to the above fungi, others that are known to cause growth/discoloration on wood which may also be used for testing include, *Alternaria* sp.; *Aspergillus brasiliensis* Varga et al. (ATCC 9642); *Aspergillus niger* van Tieghem (ATCC 6275); *Aspergillus* sp.; *Ceratocystis* sp.; *Cytospora* sp. (Pine); *Graphium* sp.; *Penicillium citrinum* Thom (ATCC 9849); *Penicillium* sp.; *Phialophora* sp.; *Trichoderma virens* (Miller et al.) von Arx, anamorph (ATCC 9645); *Trichoderma* sp.

10. Culture Media

10.1 *Agar Substrate*—For both stock culture tube and Petri dish cultures of the test fungi, use a nutrient medium: that is, malt extract agar (MEA, 2 % malt extract plus 2 % agar), potato dextrose agar (PDA, 0.4 % potato starch, 2 % dextrose plus 2 % agar), or similar commercial mixtures of MEA or PDA prepared in accordance with manufacturer instructions (for example, culturing substrate and growth conditions as defined by the ATCC). PDA stimulates sporulation in some sapstain fungi (for example, *Aureobasidium pullulans*). Sterilize the medium at 121 °C, 0.1 MPa, for 20 min.

11. Preparation of Inoculum

11.1 Maintain aseptic conditions when preparing individual and mixed spore suspensions. For laboratory experiments requiring a relatively small volume (about 100 mL) of inoculum, preparation using only the stock test tube cultures is an option. For larger volumes of inoculum, prepare from cultures grown on Petri dishes. Use only freshly prepared cultures to ensure vigor of the fungi.

NOTE 3—Before using any stock test tube culture, reinoculate new tubes to confirm growth and for future use.

11.2 For the preparation of a spore suspension, add 5 mL of sterile water to each culture tube or 10 mL to Petri dishes, and gently rub the surface of the MEA or PDA culture with a blunt glass rod to loosen the spores. Carefully aspirate the spore suspension from plate using a sterile pipet, avoiding collection of mycelial fragments or agar.

11.3 For nonsporulating cultures, obtain a mycelial suspension for use by aseptically scraping the surface mycelium off and blending it with sterile water.

³ The following numbers refer to standard strains of test fungi maintained in the American Type Collection (ATCC), P.O. Box 1549, Manassas, VA 20108, www.atcc.org.